

# **Climate and human impacts inferred from a 1500-year multi-proxy record of an alpine peat bog in the South-Eastern Alps**

# Climate and human impacts inferred from a 1500-year multi-proxy record of an alpine peat bog in the South-Eastern Alps

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## 23 1. INTRODUCTION

24 Over the last decades, human activities are, directly or indirectly, driving severe environmental  
25 changes at both global and local scales (Allamano et al., 2009; Brunetti et al., 2004; Riberio et al.,  
26 2021; Simolo et al., 2010). Global warming is one of the major drivers, causing rapid biotic  
27 changes, especially in arctic and mountain regions where temperature increase is as much as twice  
28 that of the global rate (Brunetti et al., 2009). Upward migrations of alpine and nival species  
29 (Grabherr et al., 1994), changes in communities composition, and local extinctions are just a few  
30 examples of global warming impact (Lamprecht et al., 2018; Pauli et al., 2012, 2014; Rogora et  
31 al., 2018). Global warming effects are particularly deleterious for ecosystems like peatlands, that  
32 rely on a surplus of water to act as sinks of atmospheric carbon (Riberio et al., 2021). In addition,  
33 a change of species abundance and composition in peatlands, in response to climate warming, has  
34 been shown to affect the rate of peat accumulation and decomposition, leading to an increase in  
35 the wooded area of the peatland (Dinella et al., 2021; Heijmans et al., 2008).

36 Investigating the inter-play between climatic, environmental, and biotic changes is of fundamental  
37 importance for determining the future of an ecosystem and activating the most appropriate  
38 management policies and practices (Li et al., 2020). In this respect, the reconstruction of historical  
39 environmental changes could represent an important source of information (Andrews et al., 2022;  
40 Railsback et al., 2020). To this purpose, sediment depositions represent one of the most interesting  
41 matrices to study historical climatic and environmental alterations and their driving factors (Kaal  
42 et al., 2020; Stančikaitė et al., 2019; Zhang et al., 2020).

43 Ombrotrophic bogs, i.e. peatlands dominated by peat mosses (*Sphagnum* species), are valuable  
44 climatic and biological archives. These bogs are exclusively fed by external inputs (aerosol, rain,  
45 etc.), thus providing a signal primarily related to the atmosphere and, therefore, to climate

46 variability (Damman, 1986; Rydin and Jeglum, 2006). Moreover, the anoxic conditions are  
47 fundamental for preserving organic matter: the absence of oxygen in the deepest layers reduces  
48 degradation processes. Acid-resistant organic materials such as keratinized residues of vertebrates  
49 and invertebrates, pollen, plant residues and plant macro-fossils, and testate amoebae can be  
50 therefore analyzed to establish the presence of specific taxa (Gałka et al., 2015, 2018; Kajukało et  
51 al., 2016; Lamentowicz et al., 2015; Poto et al., 2013) and assess their function in the ecosystem  
52 (Birks, 2020; Marcisz et al., 2020). Moreover, the accumulation of organic matter allows  
53 measuring peat (= organic carbon) accumulation, a function of the rate of vegetation decay  
54 (Lindsay, 1995). Variations in peatland hydrology determine changes in the rate of peat  
55 accumulation/degradation (Cristea et al., 2014; Drollinger et al., 2020), affecting the survival of  
56 living organisms such as testate amoebae (Lamentowicz et al., 2008; Mitchell et al., 2008a) and  
57 peatland trees growing on the surface (Dinella et al., 2019).

58 Therefore, by combining information on hydrology and physicochemical characteristics of  
59 peatlands with the occurrence of specific taxa, it is possible to reconstruct a sufficiently long time  
60 series of climatic and biological information to assess changes in abiotic and biotic ecosystem  
61 components. Indeed, several studies confirmed the potential of this multi-proxy approach to get an  
62 overview of peatlands dynamics and past environmental changes (Lamentowicz et al., 2010; van  
63 der Knaap et al., 2011).

64 The main targets of taxonomic identification of peatland-derived organisms are plants and testate  
65 amoebae. The former, usually identified by pollen, represent key organisms in different  
66 ecosystems and can be used as bioindicators within the studied peatland and its surroundings  
67 habitat (Rolli et al., 2015; Smith, 1994; Zurayk et al., 2001). How testate amoebae communities  
68 are assembled is linked to peatland hydrology and this information can be accordingly used as

69 indicators of the degree of aridity/humidity of the ecosystem in response to local climate  
70 (Woodland et al., 1998; Mitchell et al., 2008b). In addition, both sub-fossil and living trees on  
71 peatlands can be used as proxies of past water table variations based on their tree-ring records  
72 (Edvardsson et al., 2016; Dinella et al. 2019).

73 The most common technique for taxa identification is the morphological observation of biological  
74 residues. The success of these morphology-based techniques has been reported in many studies  
75 (Gałka et al., 2018; Lamentowicz et al., 2015; Parducci et al., 2015). However, it presents some  
76 limitations: i) it is highly dependent on the taxonomic expertise, ii) it is time-consuming; iii)  
77 especially for pollen, it is not so uncommon that the same morphological features are shared across  
78 different species.

79 The environmental DNA (eDNA) metabarcoding offers a solution to overcome these limitations  
80 (Thomsen et al., 2015). This approach has been widely applied for monitoring modern biological  
81 communities in water and soil (Edwards et al., 2018; Mayer et al., 2021; Rota et al., 2020; Sakata  
82 et al., 2020), and for the identification of plant taxa within palaeoecological archives such as lake  
83 sediments and glaciers (Alsos et al., 2018; Parducci et al., 2017). To the best of our knowledge,  
84 only one study (Parducci et al., 2015) has applied the eDNA metabarcoding approach to study  
85 plants in peatlands. Moreover, very few studies (only one as far as we know: Garcés-Pastor et al.,  
86 2019) applied eDNA metabarcoding to identify invertebrate communities, which are very  
87 abundant in peatlands and are not considered as proxies even though they have key roles and can  
88 be used as bioindicators of the ecosystem health (Gerlach et al., 2013; Paoletti et al., 1991). The  
89 present study pursues a two-fold goal. On one hand reconstructing the past climate variability of  
90 South Tyrol (Italy), while identifying possible human-induced perturbations based on the analyses  
91 of pollen and testate amoebae, and physico-chemical features of peat. On the other hand,

92 evaluating the effectiveness of the plastidial marker trnL (Taberlet et al., 1991) for the  
93 identification of plant taxa in association with the use of a portion of the mitochondrial DNA  
94 cytochrome oxidase 1 (CO1) for the classification of invertebrate taxa (Hebert et al., 2003).

## 95 **2. MATERIALS AND METHODS**

### 96 *2.1 Study site and peat coring*

97 The study site (Fig.1) is the “Biotop Wölflmoor”, an ombrotrophic peatland (= bog) located in the  
98 municipality of Nova Ponente (South Eastern Italian Alps, latitude 46,418586 – longitude  
99 11,42801), Province of Bolzano (Italy). The study area is characterized by a temperate continental  
100 climate with the highest precipitation during the summer season. The mean annual temperature is  
101 6.0°C going from a minimum of 1.1°C to a maximum of 10.6 °C, while the total rainfall is about  
102 780 mm per year (data from Nova Ponente meteorological station, period 1930-2017, Bolzano  
103 province meteorological service).

104 The peatland is included in the Natura 2000 network and represents one of the highest settings of  
105 peatlands in Europe with a minimum elevation of 1,291 m a.s.l. and a maximum elevation of 1,298  
106 m a.s.l. Its surface area is 10 hectares, and the vegetation is typical of ombrotrophic bogs with a  
107 dominance of *Sphagnum* mosses, surrounded by mountain pine (*Pinus mugo* Turra) (Alber et al.,  
108 1996; Bragazza et al., 2005). The typical vegetation at this altitude is mainly characterized by the  
109 presence of spruce (*Picea abies* (L.) H.Karst.), fir (*Abies alba* Mill.), scots pine (*Pinus sylvestris*  
110 L.) and sporadically some beech (*Fagus sylvatica* L.) (Agriculture and forestry - administration of  
111 Bolzano province).

112

113 The peat sampling, carried out during summer 2017, was performed by extracting a one-meter-  
114 long peat core using a Wardenaar peat sampler (Wardenaar, 1987). The core was then placed into  
115 a plastic tube and wrapped with a plastic foil. At the laboratory, the peat core was divided  
116 longitudinally into two halves. The first half was partitioned into slices of 1 cm so obtaining 100  
117 peat subsamples used for radiocarbon and lead dating, and the measurement of bulk density, water  
118 content,  $^{13}\text{C}$  and C:N ratio. Another subsample, taken every 4 cm, was then used for the  
119 palynological analysis and the testate amoeba analysis; the second half was further divided into 4  
120 cm-long slices for environmental DNA extraction and amplification in order to obtain a total of 23  
121 peat samples.

## 122 *2.2 Dating and geochemical proxies*

123 Peat dating was performed using  $^{14}\text{C}$  (Beta Analytic Testing Laboratory, Miami, Florida, U.S.)  
124 and  $^{210}\text{Pb}$  (Institute of Nuclear Physics Polish Academy of Science, Kraków, Poland). Eight  $^{14}\text{C}$   
125 (AMS-Accelerator Mass Spectroscopy) dates were carried out on carefully selected plant macro-  
126 fossils from 1-cm long peat slices of the profile (laboratory code: Beta; Tab. 1). Radiocarbon dates  
127 were then calibrated to receive calendar age using IntCal20 (Reimer et al., 2020) and post-bomb  
128 NH1 (Hua et al., 2021) atmospheric curve (Tab. 1). For the measurements of  $^{210}\text{Pb}$  we selected 71  
129 contiguous samples from a depth of 71–0 cm. The activity of  $^{210}\text{Pb}$  was determined as the activity  
130 of its daughter radionuclide  $^{210}\text{Po}$  (half-life 138 days), which is in radioactive equilibrium with  
131  $^{210}\text{Pb}$ . The  $^{210}\text{Po}$  activities were measured using Alpha Duo spectrometer with Ortec detectors after  
132 sample preparation following Mroz et al. (2017). Blanks and the reference material (IAEA 447  
133 moss soil) were analyzed to ensure the quality of measurements, while the age-depth relationship  
134 was estimated using the Constant Rate of Supply (CRS) model. Combining the two dating  
135 methods, we prepared the absolute chronology using i) Bayesian age-depth model based on three

136  $^{14}\text{C}$  dates calculated for a depth of 103–50 cm using the OxCal v. 4.4 software (*P\_Sequence*  
137 command:  $k=0.5$ ,  $\log_{10}(k/k_0) = 1$ ; Bronk Ramsey, 1995, 2006, 2008) applying the IntCal20 (Reimer  
138 et al., 2020) atmospheric curve as the calibration set and ii)  $^{210}\text{Pb}$  dates for a section of 50–0 cm.  
139 As water content, bulk density and C/N ratio revealed a strong difference between peat layers  
140 below and above a depth of ca. 66 cm, a boundary was applied in the  $^{14}\text{C}$ -based Bayesian age-  
141 depth model. All  $^{14}\text{C}$  dates retrieved from the section of 50–0 cm were used to validate the  $^{210}\text{Pb}$   
142 chronology. As the absolute chronology derived from the  $^{14}\text{C}$  -based Bayesian age-depth model  
143 we used  $\mu$  value (unit: cal. CE; CE-Common Era).

144 The analyses of water content, bulk density,  $^{13}\text{C}$  isotopic signature ( $\delta^{13}\text{C}$ ) and C:N ratio were  
145 conducted at the Free University of Bolzano. Water content and bulk density were calculated by  
146 measuring the wet weight and volume of each subsample and dry weight after oven drying at  $65^\circ\text{C}$ .  
147 The  $\delta^{13}\text{C}$  and C:N ratio were measured with a continuous-flow isotopic ratio mass spectrometer  
148 (CF-IRMS; Delta V Advantage, Thermo Fisher Scientific, Bremen, Germany) coupled with a  
149 CHN elemental analyzer (Flash EA 2000 Thermo Fisher Scientific, Bremen, Germany), after  
150 drying subsamples at  $105^\circ\text{C}$ .

### 151 *2.3 Pollen, testate amoebae and plant macro-fossils*

152 Palynological analysis, testate amoeba and plant macro-fossils analysis were done at the  
153 Laboratory of Climate Change Ecology of the Adam Mickiewicz University in Poznań (Poland).  
154 A total of 27 samples were prepared for palynological analyses, using the standard laboratory  
155 procedures (Berglund and Ralska-Jasiewiczowa, 1986). To remove carbonates, samples were  
156 treated with 10% hydrochloric acid. This step was followed by digestion in hot 10% potassium  
157 hydroxide (to remove humic compounds). Next, acetolysis was performed. One *Lycopodium* tablet  
158 (produced by the Lund University) was added to each sample during the laboratory procedures to



159 calculate microfossil concentration (Stockmarr, 1971). Pollen, spores, and selected non-pollen  
160 palynomorphs (NPPs) were counted under an upright microscope until the number of total pollen  
161 sum (TPS) grains in each sample reached at least 500. Sporomorphs were identified with the  
162 assistance of atlases and keys (Moore et al., 1991; Beug, 2004; van Geel and Aptroot, 2006). The  
163 percentage diagram was drawn using the TILIA Graph program (Grimm, 1991).

164 For testate amoebae analysis, peat samples were washed under 300  $\mu\text{m}$  sieves following the  
165 method described by Booth et al. (2010). Testate amoebae were analyzed under a light microscope  
166 with 200 $\times$  and 400 $\times$  magnifications, aiming at a minimum of 100 tests per sample (Payne and  
167 Mitchell, 2009). Several keys and taxonomic monographs were used to achieve the highest  
168 possible taxonomic resolution (e.g., Ogden and Hedley, 1980; Clarke, 2003; Mazei and Tsyganov,  
169 2006; Siemensma, 2021). The results of testate amoeba analysis were used for the quantitative  
170 depth-to-water table (DWT) reconstruction, which was performed in C2 software (Juggins, 2007)  
171 using the European training set (Amesbury et al., 2016).

172 Plant macro-fossils analysis was based on the accessible literature (Grosse-Brauckmann, 1986;  
173 Warner, 1990; Tobolski, 2000). The peat material was rinsed with water onto a sieve with mesh  
174 size 0.25 mm. The residue was sorted under a stereoscopic microscope under 10–100  $\times$   
175 magnification. Two randomly chosen samples from each section were examined at 200–400  $\times$   
176 magnification to estimate the volume percentage of each plant taxon.

#### 177 *2.4 Environmental DNA*

178 All laboratory procedures were carried out in the “Ancient DNA laboratory” (exclusively  
179 dedicated to ancient DNA) at Fondazione Edmund Mach – Research and Innovation Center.  
180 Sample preparation (cleaning and manual homogenization), DNA extraction and PCR

181 amplification were performed in three different rooms to avoid sample contamination. To further  
182 limit environmental contamination, the DNA extraction room was sterilized for 30 minutes using  
183 UV lamps, while all extraction steps were conducted under a biological hood and by sterilizing the  
184 equipment using bleach, ethanol, and UV for 20 minutes.

185 The DNA extraction was performed on 10 g of the sample using the DNeasy PowerMax Soil Kit  
186 (QIAGEN Inc.) and by following the manufacturer's instructions. All extraction batches included  
187 9 samples and 1 blank extraction control.

188 A short fragment of the chloroplast trnL was amplified to identify plant DNA using the primer pair  
189 c-A49325 and h-B49466 [5'-CGAAATCGGTAGACGCTACG-3' and 5'-  
190 CCATTGAGTCTCTGCACCTATC-3'] (Taberlet et al., 2007), while to identify arthropod DNA,  
191 a short fragment of the CO1 was amplified using the universal primers ZBJ-ArtF1c [5'-  
192 AGATATTGGAACWTTATATTTTATTTTGG – 3'] and ZBJ-ArtR2c [5' –  
193 WACTAATCAATTWCCAAATCCTCC – 3'] (Zeale et al., 2011), which allowed the  
194 amplification of fragments of 157 bp (reaction mix and conditions in Table S1).

195 Reagents and environmental contaminations were monitored by adding blank controls to each  
196 amplification batch consisting of 9 samples, 1 extraction control and 1 PCR blank control. Two  
197 PCR replicates were performed for each sample.

198 Both samples and blank controls were checked on the QIAxcel capillary electrophoresis, with  
199 DNA High Sensitive cartridge (Qiagen, GmbH, Hilden, Germany), and sequenced using MiSeq  
200 Reagent Kit v3 in an Illumina MiSeq platform. Only blank controls presenting a positive signal on  
201 the capillary electrophoresis were sequenced together with biological samples.

202 *2.5 Bioinformatics*

203 Raw sequences derived from trnL and CO1 markers were analyzed separately. The data were pre-  
204 processed, quality filtered, trimmed, de-noised, merged, and analyzed using QIIME2 (Bolyen et  
205 al., 2019) and DADA2 (Callahan et al., 2016). Then, the obtained Amplicon Sequence Variants  
206 (ASVs) were clustered into OTU with a cutoff of 97% similarity and classified using the  
207 VSEARCH algorithm (Rognes et al., 2016). OTUs derived from CO1 libraries were assigned using  
208 the BOLD reference database (Ratnasingham and Hebert, 2007). Plant OTUs were taxonomically  
209 classified with a custom-made reference database.

### 210 **3. RESULTS**

#### 211 *3.1 Absolute chronology and peat accumulation rate*

212 The overall trend of  $^{210}\text{Pb}$  activity versus depth displayed a progressive decline from the surface  
213 until 17 cm depth. However, there were significant departures from this simple monotonic  
214 decrease, presumably reflecting episodic variations in the rate of peat growth or decomposition.  
215 Below 17 cm,  $^{210}\text{Pb}$  unsupported activity decrease followed an approximate exponential downward  
216 trend with the increasing profile depth.

217 The age of the samples was calculated for the middle of the depth of the layers.  $^{210}\text{Pb}$  dating  
218 revealed that the peat section 0-51 cm was accumulated in  $147 \pm 42$  years. Based on the total  $^{210}\text{Pb}$   
219 inventory in the cores, atmospheric  $^{210}\text{Pb}$  fluxes were calculated as  $136 \text{ Bq m}^{-2} \text{ yr}^{-1}$ . This value  
220 agreed well with  $^{210}\text{Pb}$  flux reported by Mróz et al. (2017) for Southern Poland ( $131\text{--}160 \text{ Bq m}^{-2}$   
221  $\text{yr}^{-1}$ ,  $50^\circ\text{N}$ ) or by Vaasma et al. (2017) for North-East of Estonia ( $133 \pm 24 \text{ Bq m}^{-2} \text{ yr}^{-1}$ ,  $59^\circ\text{N}$ ) and  
222 similar  $^{210}\text{Pb}$  flux reported by Baskaran (2011), who estimated the value  $155 \pm 75 \text{ Bq m}^{-2} \text{ yr}^{-1}$  for  
223 the latitude belt  $40\text{--}50^\circ\text{N}$ .

224 The Bayesian age-depth model for a depth of 103–50 cm revealed agreement of model ( $A_{\text{model}}$ )  
225 equal to 88.3 % which is above the recommended by Bronk Ramsey (2008;  $A_{\text{model}} > 60\%$ ). This

226 section of the profile spans a period of  $550\pm 50 - 1185\pm 145$  cal. CE. The  $\sigma$  error of modelled dates  
227 ranges between 40 and 114 calibrated years (Fig.2).

228 The distinct change of age between dates at a depth of 55.5 and 50.5 cm (covering ca. 800–930  
229 calibrated years) suggests a depositional gap (hiatus) at least in this section. Moreover, at a depth  
230 of 50.5 cm the difference between  $^{210}\text{Pb}$  and  $^{14}\text{C}$  is 44–136 years, but from a depth of 45.5 cm  
231 toward the top calibrated post-bomb  $^{14}\text{C}$  dates overlap with  $^{210}\text{Pb}$  ones. Hence, the absolute  
232 chronology of the section between 55.5 and 45.5 cm is rather uncertain.

233 The section of peat below the hiatus accumulated with a rate of 0.1–0.09 cm yr<sup>-1</sup> (ca. 550–910 cal.  
234 CE; 103–66.5 cm) and ca. 0.06 cm yr<sup>-1</sup> (ca. 910–1090 cal. CE; 66.5–55.5 cm). The section above  
235 the hiatus revealed a peat accumulation rate ranging between 0.45 (ca. 1951±10 cal. CE) and 2 cm  
236 yr<sup>-1</sup> (ca. 2015 cal. CE).

### 237 *3.2 Geochemical analyses*

238 The abrupt change recorded in the peat profile at c. 50 cm was also detected by measuring the  
239 water content, bulk density,  $^{13}\text{C}$  isotope, and C:N ratio (Fig.3) and it displayed a consistent trend.

240 A large variation occurred in about 10 cm of the peat profile (ca. 1090-1952 cal. CE; 55.5 – 45.5  
241 cm), where water content increased (from 83 to 96 %) and conversely bulk density values  
242 decreased (from 0.18 to 0.03 to g/cm<sup>3</sup>). This change was detectable also in the carbon isotope  
243 measure and the C:N ratio, but to a lesser extent. The  $\delta^{13}\text{C}$  variation of bulk peat along depth can  
244 be divided into three phases. From the bottom of the core, it showed a decrease of 4.8‰ from the  
245 maximum of -20.9‰ (ca. 611 cal. CE; 96.5 cm) to the first minimum -25.7‰ (ca. 1109 cal. CE;  
246 54.5 cm), then it increased of 3.0‰ until the value of -22.7‰ (ca. 1991 cal. CE; 22.5 cm) and,  
247 finally, it reached the second minimum of -26.04‰ (ca. 2013 cal. CE; 6.5 cm) decreasing by  
248 3.34‰. The C:N ratio followed a similar pattern consisting of three stages with a slow decrease

249 from the bottom of the core, an abrupt increase in the middle part (ca. 1912-1952 cal. CE; 49.5–  
250 39.5 cm) and then a relatively stable phase until the uppermost peat layer.

### 251 3.3 Pollen analysis

252 Phase I (102.5–74.5 cm; ca. 550–720 cal. CE)

253 The period was characterized by the highest forest cover (AP: 93–97.6 %). Norway spruce (*Picea*  
254 *abies*; 15.9–33.9%), silver fir (*Abies alba*; 3.4–14.1%), common beech (*Fagus sylvatica*; 6.5–14.3  
255 %) and oak (I; 4.8–10.7 %) were the main components of local forests (Fig.4). Hop-hornbeam  
256 (*Ostrya*) and/or Oriental hornbeam (*Carpinus orientalis*), European larch (*Larix*) and hazel  
257 (*Corylus*) occurred as additional forest components. Betula trees (*Betula alba* type) and alder trees  
258 (*Alnus glutinosa* type) probably occupied the peatland outskirts and/or damp habitats in lower  
259 altitudes. High values of *Pinus sylvestris* type (23.1–30.5%) result from the proximity of *Pinus*  
260 *mugo* shrublands. Cyperaceae (1.3–3.6%), *Vaccinium* groups (0.5–3.1%), and *Sphagnum* (0.2–  
261 3.1%) sporomorphs were the most common taxa related to peatland habitat.

262 Phase II (74.5–62.5 cm; ca. 720–970 cal. CE)

263 The forest cover declined (AP: 91.4–92.8%). Among dominant taxa in the previous zone, *P. abies*  
264 (to 11.4%) and *A. alba* (to 3.9%) retreated distinctly, while a gradual increase in values of Poaceae  
265 (2.7–3%), *Plantago lanceolata* (0.6–0.8%), Cerealia type, *Secale cereale* as well as *Betula alba*  
266 type and *Corylus* took place. Continuous, but not frequent presence of *Olea* and *Vitis vinifera*  
267 pollen grains were also detected.

268 Phase III (62.5–42.5 cm; ca. 970–1958 cal. CE, with the section with uncertain chronology)

269 Deforestation affected *F. sylvatica* (decline to 2%) and *Quercus* (decline to 2%) stands the most  
270 and to a lesser degree *P. abies* and *A. alba*. *Pinus mugo* probably spread (*P. sylvestris* type: 34.3–  
271 49%). *Pinus cembra* may have appeared in the peatland vicinity. Pollen percentages of *Olea*, cf.  
272 *Phillyrea*, *Vitis vinifera* and *Humulus/Cannabis* revealed maxima in the profile (1.8, 1.3, 0.8, and  
273 1.5%, respectively), while values of Poaceae, *Plantago lanceolata*, *Artemisia*, Cerealia type and  
274 *S. cereale* (max. 0.8%) increased.

275 Phase IV (42.5–16.5 cm; ca. 1958–1987 cal. CE)

276 Decrease in AP values (77.1–84.2%) indicates continuous deforestation, which probably affected  
277 the most *F. sylvatica* and *Quercus*. Simultaneously *Ostrya* and/or *Carpinus orientalis* gradually  
278 spread (pollen type: 1.7–3.9%). At the end of Phase IV, *Betula alba* type reached its maximum  
279 (13.3%) while *Pinus mugo* (*P. sylvestris* type) retreated.

280 Phase V (16.5–0 cm; ca. 1987–2017 cal. CE)

281 Arboreal vegetation increased its cover, mainly by the expansion of *Ostrya* and/or *Carpinus*  
282 *orientalis* and partly *Pinus mugo* (*P. sylvestris* type). The decline of AP values at the top of the  
283 profile is a result of the bias related to the increase in Apiaceae values, which was probably a local  
284 event. Since ca. 1994 cal. CE a stable presence of pollen of *Ambrosia artemisiifolia* type was noted  
285 which is probably a result of the spread of invasive *Ambrosia sp.* in the region.

286 *3.4 Testate amoebae and plant macro-fossils*

287 Testate amoeba communities recorded a shift between the first and the second half of the peat core  
288 (Fig.5). Up to about 1005 cal. CE (~ 60.5 cm) the testate amoeba communities were dominated by  
289 several mixotrophic taxa: *Archerella flavum*, *Heleopera petricola*, *Heleopera sphagni* and  
290 *Hyalosphenia papilio*. During that time water table depth (DWT) at the peatland was high and

291 stable (DWT of ca. 6 cm). Between 1056 and 1954 cal. CE (ca. 57.5-44.5 cm) we recorded a shift  
292 in the relative abundance of testate amoeba species and much lower concentration of individuals  
293 in the sediment (see testate amoeba sum on Fig.5). Mixotrophic taxa decrease rapidly (except for  
294 *Heleopera petricola*), whereas small taxa (with shells smaller than 60  $\mu\text{m}$ , *Cryptodifflugia*  
295 *oviformis* and *Trinema enchelys*) and the dry indicator species *Bullinularia indica* appeared. This  
296 shift is related to rapid water table lowering (drop from ca. 6 cm to 20 cm). From 1960 cal. CE  
297 (ca. 41.5 cm) up to the top of the profile some of the mixotrophic species reappeared in the  
298 communities (*H. sphagni* and *H. papilio*) accompanied by dry indicator taxa, such as *Alabasta*  
299 *militaris*, *Nebela collaris* and *Nebela tincta*. The water table remained low in this period (mean  
300 value: 14 cm).

301 The analysis of plant macro-fossils provides insightful clues on local vegetation changes. The  
302 diagram (Fig.6) can be divided into two sections. The bottom part of the core till 72.5 cm (ca. 841  
303 cal. CE) is represented by the domination of *Sphagnum fuscum/rubellum*, Ericaceae root and  
304 herbaceous plant remain with the addition of *Sphagnum papillosum*, *Sphagnum cuspidatum* and  
305 *Andromeda polifolia* that occurs in form of leaves and seed on depth 90-87 cm. Also, Pine and  
306 *Rhynchospora* seeds were recorded in this section. The horizon 65.5-50.5 cm (ca. 919-1185 cal.  
307 CE) consisted of a critical threshold in the peatland development, the strongest change at 61.5 cm  
308 (ca. 988 cal. CE) represented by the peak of the unidentified organic matter being simultaneously  
309 the most decomposed part. Above 61 cm the plant communities were dominated by *Sphagnum*  
310 *magellanicum* and *Sphagnum papillosum*, while herbaceous plants disappeared at a depth of 52.5  
311 cm (ca. 1148 cal. CE). *Rhynchospora* seeds were recorded until the depth 41 cm (ca. 1960 cal.  
312 CE). This section consisted of a low percentage of Ericaceae roots.

313 *3.5 Environmental DNA taxonomic classification*

314 The analysis of plant eDNA provides a similar trend as that from all the other proxies. The highest  
315 abundance is distributed between three main families: Sphagnaceae, Cyperaceae and Ericaceae  
316 (Fig.7). Sphagnaceae and Ericaceae are dominant between 45.5 and 36.5 cm depth (ca. 1952-1968  
317 cal. CE) but present a strong decrease of the abundances in superficial and deep samples,  
318 particularly for Ericaceae as already presented by the macro-fossil analysis. Cyperaceae followed  
319 an opposite trend, missing completely between 45.5–36.5 cm and dominating the superficial and  
320 deep samples. All other taxa are much less represented; nevertheless, they appear to present  
321 slightly more variability in the most superficial samples than in the intermediate and deepest ones.  
322 Finally, Betulaceae were found only at a depth of 63.4 cm (ca. 954 cal. CE), whereas they were  
323 found in almost all samples with the pollen analysis.

324 The analysis of the mt DNA CO1 revealed the high abundance of the Adinetidae family (Fig.8a),  
325 which belongs to the Rotifera phylum. Among all the other taxa, belonging to the Arthropoda  
326 phylum (Fig.8b), Carabidae and Blastobasidae represented the most abundant families, with the  
327 first one occurring only in deep samples, and the second one in some superficial and deep samples,  
328 showing a similar pattern to the other proxies before and after the hiatus. The other families were  
329 recorded in traces, but they seem to have been influenced by the environmental conditions since  
330 the Gnaphosidae family, together with Tabanidae and Cecidomyiidae, appeared only in samples  
331 corresponding to the drier period.

#### 332 **4. DISCUSSION**

333 Peat bogs from mountain areas are of particular interest for reconstructing climatic and associated  
334 ecological changes through time. Peat bogs can be precisely dated, and a series of well-established  
335 proxies offers the opportunity to evaluate changes in abiotic and biotic components (Lamentowicz  
336 et al., 2010) by integrating physicochemical, hydrological, and taxonomical information. The latter



337 is, however, sometimes difficult, and time-consuming to obtain, especially when targeting both  
338 fauna and flora. In this study of the Wölflmoor peat bog, situated in the Southern Alps (South  
339 Tyrol, Italy), we aimed at, on one hand, reconstructing climatic and associated ecological changes  
340 with established proxies and methodologies, and, on the other hand, assessing the effectiveness of  
341 eDNA metabarcoding for taxonomic identification of plant and invertebrate taxa.

#### 342 *4.1 Climatic and ecological changes*

343 From our analysis, a substantial change in peatland conditions before and after the hiatus occurred.  
344 Before the hiatus, between ca. 920-1186 cal. CE (65.5-50.5 cm) water content and bulk density  
345 decrease and increase, respectively, highlighting a period of important changes in peatland  
346 conditions (Fig.3). The clear trend in the water content and bulk density may be interpreted as a  
347 shift from moist to dry environment, which persisted for almost a century (ca. 1090-1186 cal. CE;  
348 55.5-50.5 cm). As already recorded in other peatlands across Europe (Lamentowicz et al., 2008,  
349 2009) and other natural archives (Esper et al., 2002; Tiljander et al., 2003), the dry phase may be  
350 explained by the Medieval Warm Period (WMP), which was characterized by temperatures  
351 comparable to the current ones (Bradley et al. 2003; Crowley and Lowery, 2000; Goosse et al.  
352 2006).

353 The other proxies also highlighted drier conditions and a more mineral soil substrate shift in the  
354 same time window. Plant macro-fossils (Fig.6) revealed a dry phase at ca 60.5 cm (ca. 1005 cal.  
355 CE; critical transition) described by the decomposed peat and indicate a disturbance that started at  
356 ca. 65.5 cm (920 cal. CE) and intensified at ca. 60.5 cm (1005 cal. CE). The site recovered from  
357 the disturbance until 55.5 cm, when peat initiated again a very rapid accumulation. The testate  
358 amoeba communities (Fig.5), especially in terms of individual concentration (lower testate amoeba  
359 sums analyzed in this period) and testate amoeba functional trait composition, showed a significant

360 drop in the relative abundance of mixotrophic species during the critical period. A high abundance  
361 of mixotrophs is linked to high water tables and an open landscape that allows high light intensity  
362 on the surface of the bog (Heal, 1964; Payne et al., 2016; Marcisz et al., 2016; Creevy et al., 2018).  
363 Water table lowering and shading due to the overgrowth of the peatland with trees, as confirmed  
364 by the presence of wood in this phase, could harm phototrophic metabolism, leading to the  
365 disappearance of mixotrophs from testate amoeba communities (Jassey et al., 2015; Payne et al.,  
366 2016; Marcisz et al., 2020) and promoting smaller taxa, usually indicator of dry conditions  
367 (Marcisz et al., 2016).

368 At this stage, also the pollen stratigraphy confirmed important vegetational changes (Fig.4).  
369 During Phase I (102.5–74.5 cm; ca. 550–720 cal. CE), the high forest cover dominated by *Picea*  
370 *abies*, *Abies alba* and *Fagus sylvatica* is related to a wet environment (Lamentowicz et al., 2008,  
371 2015; Gałka et al., 2015), presumably with low human pressure. Phase II, on the other hand, seems  
372 to have been a scenario of a forest decline, characterized by the retreat of *Picea abies* and *Abies*  
373 *alba* and an increase of Poaceae, Cerealia type, *Secale cereale* as well as *Betula* and *Corylus* (Phase  
374 II: 74.5–62.5 cm; ca. 720–970 cal. CE), which may suggest adaptation of deforested areas to  
375 cultivated land and pastures as well as colonization of some deforested abandoned lands by birch  
376 and hazel (within forest gaps). Subsequent phase III (62.5–42.5 cm; ca. 970–1958 cal. CE)  
377 revealed a progression in the deforestation process with a further increase of Poaceae, *Plantago*  
378 *lanceolata*, Cerealia and *Secale cereale* values, indicating a high human pressure. At the same  
379 time, an increase in Cyperaceae percentages compared to *Sphagnum* suggests a shift to drier  
380 environmental conditions.

381 After the mentioned section, the age-depth model showed the presence of a hiatus which is a  
382 consequence of a disturbance that completely transformed the bog vegetation. Similar transitions

383 in time were connected with a human impact and described from the Polish Carpathians (peat  
384 cutting) (Kołaczek et al., 2018) as well as Pomerania (unknown origin) (Lamentowicz et al., 2008).  
385 Judging from the C:N, bulk density, and testate amoeba data this depositional gap is highly related  
386 to strong and prolonged desiccation of peat, which caused faster decomposition of the topmost  
387 layer. However, due to the high human pressure, mechanical removal of the upper peat layer should  
388 be also taken into consideration as a cause of such distinct hiatus at a depth of ca. 50 cm. Results  
389 from layers after the hiatus were compared with data produced by the monitoring station, which  
390 identified dry periods in 1945, in the early 1970s, around 2005, and in the last five years (Fig.9),  
391 while short episodes of moist conditions occurred in 1979 and before the beginning of 2000. These  
392 observations are confirmed by the testate amoeba communities, that generally followed the  
393 precipitation pattern. From 1960 cal. CE mixotrophic species reappeared inside the communities,  
394 which combined with the presence of dry indicator taxa corroborate the alternation of dry and wet  
395 periods. The pollen data (Phases IV and V) highlight continuous deforestation, with an increase of  
396 *Betula alba* type ca. 1987 cal. CE, which may indicate birch colonization ca. 1984 cal. CE on  
397 abandoned open-lands which were mainly occupied by grasslands.

#### 398 *4.2 eDNA metabarcoding of plant and invertebrate taxa*

399 As the other proxies, eDNA metabarcoding revealed an abrupt change in the peatland conditions  
400 dominated by the disappearance of Cyperaceae around the depth of 45.5 cm (ca. 1925 cal. CE)  
401 together with the appearance of Ericaceae. As shown by Fig.7, the identified wet periods are  
402 dominated by Cyperaceae, a family of vascular plants related to wet environments (Gałka et al.,  
403 2018), while the abundance of Sphagnaceae and Ericaceae, more adapted to drier conditions  
404 (Gałka et al., 2018), decrease drastically. In contrast, the dry period is characterized by a high  
405 abundance of the last two families, while Cyperaceae completely disappeared.

406 Because of their strong adaptation to peatland conditions, Sphagnaceae, Ericaceae and Cyperaceae  
407 were certainly present in higher proportions (Farrick and Price, 2009; Svensson, 1986; Vitt, 2006).  
408 Therefore, DNA from these families could have been preferentially amplified, causing an under-  
409 representation of all the other species (Fulton, 2012).

410 Among the metazoans, the most abundant taxa were assigned to the Rotifera phylum (Fig.8a). This  
411 phylum is known to include organisms able of coping with the most diverse environmental  
412 conditions (Arora, 1966; Bērziņš and Pejler, 1989). This ability allows them to inhabit the deepest  
413 layers of peatlands characterized by low oxygen concentrations, which usually precludes the  
414 survival of many other organisms. For this reason the under-representation of many taxa can be  
415 related to the preferential amplification of DNA from modern organisms belonging to Rotifera  
416 (Garcés-Pastor et al., 2019). Nevertheless, excluding rotifers (Fig.8b), two families are the most  
417 common among all samples: Carabidae and Blastobasidae. The presence of the first family was  
418 established only for the deepest samples. This finding supports the identification of a shift from  
419 wet to dry conditions, as this family was found to be related to deciduous forests (Calatayud et al.,  
420 2016). Therefore, the presence of forest species around the peatland favored the establishment of  
421 Carabidae, which then was not detected with the increase of temperatures at the beginning of the  
422 dry period (Fig.8b). The second family belongs to the order of Lepidoptera. This order represents  
423 a group of organisms spread worldwide and in all terrestrial ecosystems (Lopez-Vaamonde et al.,  
424 2010). The occurrence of the Blastobasidae family is not indicative of specific environmental  
425 conditions, but according to Spitzer et al. (1993) Lepidoptera living in non-disturbed environments  
426 are specialized taxa that may be damaged by any source of the disturbance. According to Fig.8b,  
427 this family was detected in samples related to periods long before and long after the event that  
428 caused the hiatus. Therefore, it could be hypothesized that this event, in addition to causing a loss

429 of material from the peatland, resulted in a disturbance of these taxa, which were able to recover  
430 after a few years.

431 Despite their presence in traces, also the other families seems to corroborate the shift in the  
432 environmental conditions; in fact, Gnaphosidae is a family of organisms with a predilection for  
433 drier conditions (Gajdoš et al., 2016), while the Culicidae family (more adapted to wet habitat  
434 (Costa et al., 2010)) was replaced by Tabanidae, a family inside the same order of Culicidae, but  
435 with a preference versus drier environmental conditions (Herczeg et al., 2015).

#### 436 *4.3 Taxonomic identification: comparison between eDNA Metabarcoding and morphology*

437 Pollen and eDNA confirmed trends shown by the physicochemical analyses. However, it seems  
438 that the results of the eDNA metabarcoding are somehow distorted due to the signal saturation by  
439 modern and/or highly abundant taxa. This determined this method to be less effective than the  
440 morphological taxonomic identification of pollen and not fully explanatory for arthropods.

441 The reasons for this performance could be explained by the different amounts of material needed  
442 by the approaches to establish the presence of a taxon inside samples. For the morphological  
443 identification, the presence of a single pollen grain is sufficient, while for the eDNA the amount  
444 of starting material must be sufficient to get several reads able to pass all the filtering steps.  
445 However, this goal can be hampered by several factors: the under-representation of taxa due to the  
446 preferential amplification of abundant taxa (Shirazi et al., 2021), the accuracy of the reference  
447 databases (i.e. ideally, they should include reference sequences for each OTU in our dataset)  
448 (Schenekar et al., 2020), the degeneration of primers (i.e. having degenerate primers able to  
449 amplify DNA sequences from species also quite distant from each other) (Elbrecht et al., 2019),  
450 and the variability of the selected markers within target species (Meusnier et al., 2008; Taberlet et  
451 al., 1991).

452 It is important to recall that, insofar, very few eDNA studies (Garcés-Pastor et al., 2019; Parducci  
453 et al., 2015) have been performed on peat bogs. For this reason, we argue that eDNA  
454 metabarcoding is expected to be significantly improved. For both plants and arthropods, reference  
455 databases are becoming more comprehensive year by year, and other sets of primers (Chen et al.,  
456 2010; CBOL Plant Working Group: Hollingsworth et al. 2009; Elbrecht et al., 2019; Hadziavdic  
457 et al., 2014) can be used in place of or in support of those used in this study in order to increase  
458 the range of taxa identified at high taxonomic levels.

459 For plants, eDNA and pollen can provide complementary information able to improve the  
460 classification of taxa identified with the morphological approach alone.

461 About arthropods, they have never been investigated for historical reconstructions, probably due  
462 to the difficulties in the taxonomic assignment with the morphological identification (Thomsen et  
463 al., 2015). These difficulties can be caused by the similar appearances of some species or because  
464 morphological keys can be useful only for specific genders or life stages.

465 The eDNA can overcome these limitations, allowing the inclusion of arthropods and other  
466 invertebrates in studies based on the historical reconstruction of biodiversity since they play a  
467 fundamental role inside lots of ecosystems and can be used as bioindicators of soil quality  
468 (Hodkinson and Jackson, 2005; Maleque et al., 2009; Nakamura et al., 2007; Paoletti, 1999;  
469 Seastedt and Crossley, 1984).

## 470 **5. CONCLUSIONS**

471 The first 1-m long peat core extracted from the Wölflmoor bog in South Tyrol (Italy) is more than  
472 a millennium old, suggesting a long history of this peatland during the Holocene. The stratigraphy  
473 showed a consistent difference in the composition and structure of the material along the peat core  
474 and, consequently, the peat dating revealed a lack of material halfway through the sample, which

475 determined an interruption of about 600 years. This may suggest that the Wölflmoor bog was  
476 exploited between the end of the 19th century and the beginning of the 20th. Proxies from  
477 Wölflmoor bog showed converging trends over time with visible differences in recorded values  
478 between the two portions of the peat core. Pollen, water content, bulk density, and testate amoebae  
479 indicate a shift from moisture to warmer, drier conditions between 920 and 1186 cal. CE, possibly  
480 attributed to the initial stage of the Medieval Warm Period, while the increase in Poaceae, Secale  
481 cereale and other Cerealia type highlight a high human pressure started ca. 720 cal. CE. The eDNA  
482 metabarcoding analyses confirmed this trend, showing the coherence with climatic proxies already  
483 widely used in climate reconstruction studies.

484 In recent decades, the comparison of peatland proxies allowed detecting an alternation of dry to  
485 wet period, particularly warmer and drier conditions during the last years.

486 Overall, eDNA metabarcoding was less effective than pollen morphological identification, while  
487 for the Arthropoda the signal was likely saturated by living organisms, showing that this technique  
488 needs further methodological developments. Further studies are foreseen to explore the full  
489 potential of this technique in peatland archives studies to produce data that can effectively  
490 complement those derived by morphological analyses.

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493 This study is dedicated to the memory of Giustino Tonon.

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